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Occurrence and unlimited spread of *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) in Tanzania: A potential economic threat to the citrus industry

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ABSTRACT

Citrus production is a cornerstone of Tanzania's agricultural sector, supporting economic development and food security. However, the industry faces a critical threat from the Asian citrus psyllid (*Diaphorina citri* Kuwayama), the primary vector of the destructive Huanglongbing (HLB) disease. This study investigates the distribution, abundance, and environmental drivers of *D. citri* and the African citrus triozid (*Trioza erytreae*) in major citrus-growing regions of Tanzania. Surveys conducted between March and May 2023 revealed the presence of *D. citri* in the Tanga and Morogoro regions, including the first documented occurrence in the Tanga region. Conversely, *T. erytrae* was confined to the Arusha region. Altitude emerged as a significant factor influencing psyllid abundance. *D. citri* populations decreased with increasing altitude, while *T. erytrae* exhibited the opposite trend. Temperature also showed correlations with psyllid populations, though additional research is needed to fully elucidate these relationships. Citrus variety significantly influenced *D. citri*, offering potential sources of resistance. The intensity of new flush growth was strongly correlated with psyllid abundance. These findings highlight the urgent need for region-specific and altitude-tailored citrus pest management strategies. The identification of potentially HLB-resistant citrus varieties provides a promising avenue for breeding programs. This research offers critical insights to protect Tanzania's citrus industry and ensure the sustainable production of this vital economic crop.

Keywords: Citrus, Citrus greening, Diaphorina citri Kuwayama, Huanglongbing, Trioza erytreae, Distribution, Altitude, Tanzania

RÉSUMÉ

Présence et propagation illimitée de *Diaphorina citri* Kuwayama (Hemiptera : Liviidae) en Tanzanie : une menace économique potentielle pour l'industrie des agrumes

La production d'agrumes est un pilier du secteur agricole de la Tanzanie, soutenant le développement économique et la sécurité alimentaire. Cependant, l'industrie fait face à une menace critique du psylle asiatique des agrumes (Diaphorina citri Kuwayama), le principal vecteur de la maladie destructrice du Huanglongbing (HLB). Cette étude examine la distribution, l'abondance et les facteurs environnementaux qui influencent D. citri et le trioze africain des agrumes (Trioza erytreae) dans les principales régions productrices d'agrumes de la Tanzanie. Des enquêtes menées entre mars et mai 2023 ont révélé la présence de D. citri dans les régions de Tanga et de Morogoro, y compris la première occurrence documentée dans la région de Tanga. Au contraire, T. erytrae était confiné à la région d'Arusha. L'altitude s'est révélée être un facteur significatif influençant l'abondance des psylles. Les populations de D. citri diminuaient avec l'augmentation de l'altitude, tandis que T. erytrae présentait la tendance inverse. La température a également montré des corrélations avec les populations de psylles, bien que des recherches supplémentaires soient nécessaires pour élucider pleinement ces relations. La variété d'agrumes a influencé de manière significative l'abondance de D. citri, avec la Late Valencia (Kitenesi) étant la plus sensible. Plus important encore, plusieurs variétés n'ont montré aucune présence de D. citri, offrant des sources potentielles de résistance. L'intensité de la croissance des nouvelles pousses était fortement corrélée à l'abondance des psylles. Ces résultats soulignent la nécessité urgente de stratégies de gestion des ravageurs des agrumes spécifiques à la région et adaptées à l'altitude. L'identification de variétés d'agrumes potentiellement résistantes au HLB offre une voie prometteuse pour les programmes de sélection. Cette recherche offre des perspectives critiques pour protéger l'industrie des agrumes de la Tanzanie et assurer la production durable de cette culture économique vitale.

Mots clés : Agrumes, Verdissement des agrumes, Diaphorina citri Kuwayama, Huanglongbing, Trioza erytreae, Distribution, Altitude, Tanzanie

INTRODUCTION

Citrus fruits are a major global crop, valued for their nutritional benefits and economic importance (FAO,

2021). Their high content of phytonutrients and bioactive compounds promotes health and boosts immunity (Liu et

al., 2022). The demand for citrus fruits, especially oranges and lemons, has surged during the COVID-19 pandemic as consumers aim to increase their vitamin C intake (Andrade-Cuvi et al., 2023; Moreb et al., 2021). According to Reportlinker (2023), increased demand from both domestic and foreign markets will drive global citrus production to over 160 million tons by 2027. Brazil, China, the United States, India, and Mexico are among the major citrus-producing countries (FAO, 2022). In Africa, citrus cultivation plays a significant role in food security, employment, and export revenue. Within East Africa, Tanzania, Kenya, and Uganda play significant roles as key citrus producers (FAO, 2021). Tanzania's citrus production is steadily increasing, primarily in regions like Morogoro, Tanga, Kilimanjaro, and Coast (ESRF, 2021). Citrus farming provides direct and indirect employment opportunities, supporting the livelihoods of many rural communities and contributing to the country's agricultural and economic growth (URT, 2021). Despite its potential, the citrus industry faces significant challenges, including limited access to highquality planting materials, poor post-harvest practices, and inadequate infrastructure (Aminu et al., 2020). Furthermore, insect pests and diseases pose a constant threat to citrus production (Makorere, 2014). Among these pests, citrus psyllids are a major concern due to their ability to transmit the incurable Huanglongbing (HLB) disease (Bove, 2006). HLB, caused by bacterial pathogens (Candidatus Liberibacter spp.), is a devastating citrus disease with no known cure (Garnier et al., 1984a, 1984b). The disease has resulted in significant economic losses in citrus-growing regions worldwide (Singerman & Useche, 2019). Two insect vectors-the African citrus triozid (Trioza ervtreae) and the more destructive Asian citrus psyllid (Diaphorina citri Kuwayama)-transmit the pathogens (Graça & Korsten, 2004). The global movement of people and goods has helped the spread of HLB and its carriers, posing a significant threat to citrus production worldwide (Gottwald et al., 2007). In recent years, the spread of Diaphorina citri Kuwayama in Tanzania has increased the risk of HLB becoming widely established in the country and across Africa. Diaphorina citri Kuwayama was first detected in Eastern Africa during surveys conducted in Tanzania in 2014 and 2015 (Shimwela et al., 2016). Predictive models indicate that large areas of coastal Tanzania provide favorable conditions for the establishment of D. citri (Rwomushana et al., 2017). The significant losses incurred in other affected regions show that the potential economic impact of HLB on the citrus industry for Tanzania could be severe (de Oliveira et al., 2013). Efficient monitoring of D. citri is crucial for early detection, allowing prompt intervention and the use of customized pest control methods to protect the citrus industry (Grafton-Cardwell et al., 2013; Rogers & Stansly, 2007). However, there is a significant lack of knowledge about the current distribution and abundance of D. citri in Tanzania's citrusgrowing regions. This study aims to fill this gap by conducting systematic assessments to explore the

presence and spread of *D. citri* in key citrus production areas of Tanzania. The results will provide crucial insights, such as *D. citri* distribution patterns, population dynamics, and hotspot areas, which will assist citrus stakeholders in developing targeted pest control strategies to safeguard Tanzania's citrus sector.

Materials and methods

Study site

The study was carried out in four discrete regions of Tanzania. These are Ruvuma, located at coordinates - 10.7000 latitude and 36.2333 longitude; Arusha, situated at coordinates -3.3667 latitude and 36.6833 longitude; Morogoro, positioned at coordinates -6.8210 latitude and 37.6612 longitude; and Tanga, found at coordinates - 5.0689 latitude and 39.0988 longitude. The selection of these regions was based on their citrus production records and the different altitudes they have, as altitude is a crucial factor that affects the distribution and diversity of citrus psyllids (Jenkins *et al.*, 2015). The chosen regions encompass a variety of elevations: Tanga for low elevations (300 m), Morogoro and Ruvuma for moderate elevations (ranging from 300 m to 900 m), and Arusha for high elevations (exceeding 900 m).

Surveys and sample collection

Citrus surveys were conducted from March to May 2023. One district was chosen from each region based on citrus production records and farm availability. Ten citrus farms were randomly selected per district to ensure a representative sample. GPS coordinates (latitude, longitude) and altitude were recorded for each farm. In orchards smaller than 0.5 hectares, all citrus trees were assessed. In larger orchards, 10 trees were selected at regular intervals. On each tree, one new shoot was randomly selected from each of the four cardinal points and examined for citrus psyllids. Special attention was paid to leaf galls indicative of *Trioza erytreae*, particularly at higher altitudes. Collected psyllids were preserved in ethanol for identification and analysis.

Psyllid Identification

Initial identification of psyllids involved using a hand lens. Psyllids with a mottled brown or greyish-brown body, light brown head, and probing at a steep angle were identified as *Diaphorina citri* Kuwayama (Figure 2). Meanwhile, probing at a less steep angle, black-headed psyllids with transparent wings were identified as *T. erytrae* (Figure 3), per Shimwela *et al.* (2016). Nymphs with a white, waxy secretion and light-yellow colour with red eyes were identified as *Diaphorina* sp. Conversely, *T. erytrae* nymphs had brown eyes and, sometimes, a pair of spots on the base of the abdomen in the late stages. *Trioza erytreae* nymphs were generally found in puckered galls on the leaves.



Table 1. Description of selected products used in the study

Psyllid detection and identification

Total DNA was extracted from psyllid specimens using the CTAB protocol. DNA concentration and purity were assessed using a Nanodrop spectrophotometer. For species confirmation, a portion of the mitochondrial cytochrome oxidase I (mtCOI) gene was amplified by conventional PCR using the DCITRI COI primers (Boykin *et al.*, 2012) (Table 1). The PCR protocol was as follows:

Reaction mixture: (10 μ L total) 1x PCR buffer, 0.2 mM

dNTPs, 1 µL DCITRI COI primers.

Cycling conditions: 94°C/3 min (initial denaturation); 35 cycles of [94°C/30 sec, 60°C/30 sec, 72°C/1 min]; 72°C/10 min (final extension).

PCR products were visualized on a 1.2% agarose gel (120V, 1.5 hrs) stained with ethidium bromide, using a 1000bp ladder for size comparison. This yielded an expected 834-bp fragment for *Diaphorina citri Kuwayama* (Fujiwara *et al.*, 2017).

Table 2. Set of primers used to detect DNA on Diaphorina citri Kuwayan	<i>ıma</i> Kuwayama
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S/N	Coding region	Primer Sequence		PCR product size
1	mtCOI	DCITRI	COI-L	834 bp
		(5'-AGGAGGTGGAGACCCAATCT-3')		
		DCITRI	COI-R	
		(5'-TCAATT GGGGGGAGAGTTTTG-3')		

Data analysis

Spatial analysis and mapping of citrus farm distribution were performed using QGIS (version 3.30.2). Descriptive statistics (mean, median, standard deviation, range) were calculated for variables including altitude, temperature, and psyllid abundance. Correlation and linear regression analyses were conducted in R (version 4.3.1) to examine relationships between altitude, temperature, and the abundance of *D. citri* and *T. erytrae*.

RESULTS

Psyllid visual observations

Psyllids with a mottled brown or greyish-brown body, light brown head, and probing at a steep angle were identified as *Diaphorina citri* Kuwayama (Figure 2). Meanwhile, probing at a less steep angle, black-headed psyllids with transparent wings were identified as *T. erytrae* (Figure 3), as per Shimwela *et al.*, 2016. Nymphs with a white, waxy secretion and light-yellow colour with red eyes were identified as *Diaphorina* sp. Conversely, *T. erytrae* nymphs had brown eyes and, sometimes, a pair of spots on the base of the abdomen in ate stages. *Trioza erytreae* nymphs were generally found in puckered galls on the leaves.



Figure 2. (a) Adult *Diaphorina citri* Kuwayama, (b) and (c) Adult *Diaphorina citri* Kuwayama and its nymphs, (d) The nymphs of *Diaphorina citri* Kuwayama



Figure 3. (a) and (b) The leaf galls induced by *Trioza erytreae* on citrus shoots, (c) and (d) *Trioza erytreae* de Guercio with its eggs.

Psyllid molecular identification

Polymerase Chain Reaction (PCR) and agarose gel electrophoresis were utilised to identify *D. citri* in several samples. A 1.2% agarose gel ran at 120V for 1.5 hours. Post the run, ethidium bromide was applied to the gel to enable the visualisation of DNA bands. The specific primers used in the PCR reaction were DCITRI-COI-L and DCITRI-COI-R, specific to *D. citri*. The samples tested were TMF 6-6, TMF 4-1, TMF 6-2, MMF 8-10,

MMF 1-1, and TMF 1-0. A molecular weight marker (M) and a negative control (NC) were included in the gel in addition to the samples. The target band was of size 834bp and was visible in the lane labelled "M". This indicated the presence of *D. citri* DNA in those samples (Figure 4).



Figure 4. Molecular detection of *Diaphorina citri* Kuwayama.

The distribution of citrus psyllids

This study revealed how the occurrence of *D. citri* and *T. erytrae* in Tanzania varies across different regions. The presence of *D. citri* was limited to the Tanga and Morogoro regions, where it was present in 2 out of 10 sites in Tanga and 5 out of 10 sites in Morogoro (as seen in Table 2). Interestingly, none of the sites in Ruvuma had any *Diaphorina citri* Kuwayama. Conversely, *T. erytrae* was observed in 4 out of 10 sites in Arusha. It was found that the distribution of these pests is region-specific and varies across different areas, as demonstrated by this study (Figure 5).



Figure 5. Map of Tanzania showing locations where psyllids surveys were conducted. Blue points: locations where no psyllids were collected; black points: locations where African citrus psyllids (*Trioza erytreae* (del Guercio, 1918)) were observed; red points: locations where Asian citrus psyllids (*Diaphorina citri* Kuwayama) were observed.

Region	Farm number	Ward	Latitude	Longitude	Altitude	Variety	Psyllids Specie	HLB Symptom	Intensity of new flushes
Ruvuma	1	Nakayaya	-10.9879935	37.296125	513.5	Bitijuma	None	Present	High
Ruvuma	2	Nanjoka	-11.0601341	37.337506	664.7	Local variety	None	Absent	None
Ruvuma	3	Nanjoka	-11.0555476	37.33181	660.3	Local variety	None	Absent	Moderate
Ruvuma	4	Kangomba	-10.9797399	37.291018	531.6	Local variety	None	Present	Low
Ruvuma	5	Kangomba	-10.9686599	37.284199	533.8	Local variety	None	Present	Low
Ruvuma	6	Temeke	-10.9412059	37.392876	595.7	Local variety	None	Present	Low
Ruvuma	7	Temeke	-10.9428987	37.392738	593.6	Local variety	None	Absent	Low
Ruvuma	8	Temeke	-10.9447728	37.391446	592.6	Local variety	None	Absent	Low
Ruvuma	9	Temeke	-10.9362276	37.39519	606.3	Local variety	None	Present	Low
Ruvuma	10	Kangomba	-10.9819636	37.290017	548.4	Local variety Valencia (also	None	Absent	Low
Arusha	1	Engutoto	-3.2959374	36.449145	1554.5	known as Msasa)	Trioza erytreae	Absent	Low
Arusha	2	Engutoto	-3.3252071	36.45144	1407.1	Local variety	Trioza erytreae	Absent	Low
Arusha	3	Engutoto	-3.3260832	36.471876	1486.1	Valencia (Msasa)	None	Absent	None
Arusha	4	Engutoto	-3.3208224	36.448906	1435.8	Valencia (Msasa)	None	Absent	Moderate
Arusha	5	Engutoto	-3.3136255	36.446299	1454.4	Local variety Washington navel	None	Absent	Low
Arusha	6	Mto Wa Mbu	-3.37901	35.851073	950.9	(Kitovu)	Trioza erytreae	Present	Moderate
Arusha	7	Mto Wa Mbu	-3.3885351	35.849249	959.8	Local variety	None	Absent	Low
Arusha	8	Mto Wa Mbu	-3.3908645	35.854098	952.0	Valencia (Msasa)	Trioza erytreae	Present	High
Arusha	9	Mto Wa Mbu	-3.3866238	35.853506	941.2	Local variety	None	Absent	Low
Arusha	10	Mto Wa Mbu	-3.3701142	35.846101	961.1	Local variety	None	Absent	Low
Tanga	1	Tingeni	-5.1932635	38.74399	197.4	Valencia (Msasa) Late Valencia	None	Present	Low
Tanga	2	Tingeni	-5.1977065	38.759435	208.0	(Kitenesi) Late Valencia	None	Present	Low
Tanga	3	Tingeni	-5.202348	38.769081	179.7	(Kitenesi) Late Valencia	None Diaphorina citri	Absent	Low
Tanga	4	Tingeni	-5 183548	38 773697	161.8	(Kitenesi)	Kuwayama	Present	High
Tanga	5	Kwabadi	-5 3243437	38 743933	228.0	Valencia (Msasa)	None	Absent	High
Tonco	6	Vwahadi	5.5245457	20 746496	101.2	Late Valencia	Diaphorina citri	Dresent	High
Tanga	0	Kwabadi Kasala di	-5.5545475	38.740480	191.2	(Kitenesi)	Kuwayama	Present	High
Tanga	7	Kwabadi	-5.3230697	38./8/10/	204.6	Late Valencia	None	Present	Low
Tanga	8	Mtindiro	-5.2688045	38.802039	177.5	(Kitenesi)	None	Absent	Low
Tanga	9	Mtindiro	-5.2649714	38.792234	209.4	Valencia (Msasa)	None	Absent	High
Tanga	10	Mtindiro	-5.2482176	38.768496	238.8	Valencia (Msasa)	None Diaphorina citri	Present	Moderate
Morogoro	1	Konde	-7.0582139	37.771343	258.6	Valencia (Msasa)	Kuwayama Diaphorina citri	Absent	High
Morogoro	2	Konde	-7.0494275	37.765264	257.3	Valencia (Msasa)	Kuwayama	Present	Moderate
Morogoro	3	Tawa	-7.0525402	37.759583	263.9	Valencia (Msasa)	None Diaphorina citri	Absent	Low
Morogoro	4	Tawa	-7.0447249	37.763966	270.7	Matombo sweet	Kuwayama Diaphorina citri	Absent	Low
Morogoro	5	Konde	-7.0562604	37.762008	268.0	Matombo sweet	Kuwayama	Absent	Low
Morogoro	6	Konde	-7.0674187	37.766729	326.3	Jaffa (Shamoti)	None	Present	Low
Morogoro	7	Konde	-7.0581612	37.768089	250.2	Matombo sweet	None Diaphorina citri	Absent	Moderate
Morogoro	8	Konde	-7.0567359	37,777076	293.3	Matombo Sweet	Kuwavama	Absent	Moderate
Morogoro	9	Mlimani	-6.8410166	37.667829	498.0	Valencia (Msasa) Mediterranean	None	Present	Moderate
Morogoro	10	Mlimani	-6.8428905	37.662237	495.4	(Nairobi)	None	Present	Moderate

Psyllid abundance

The summary statistics revealed that the abundance of Diaphorina citri Kuwayama adults varied significantly across the Arusha, Morogoro, Ruvuma, and Tanga regions (p = 0.001). The mean and maximum number of Diaphorina citri Kuwayama adults were highest in Tanga, which had 3.45 per region and 86 per farm, respectively. Conversely, Arusha and Ruvuma had no Diaphorina citri Kuwayama adults in the sampled areas. The standard deviation was also highest in Tanga (12.19), indicating a high variability in Diaphorina citri Kuwayama adults' distribution. A similar pattern was observed for Diaphorina citri Kuwayama nymphs, with Tanga having the highest mean (0.5 per region) and maximum (4 per farm) number of Diaphorina citri Kuwayama nymphs, while the other regions had none (Figure 6). The abundance of *T. erytrae* adults was low across all regions except for Arusha, which had a mean of 0.153 and a maximum of 9 T. erytrae adults. The range and standard deviation of 9 and 1.172, respectively were also highest in Arusha, suggesting a skewed distribution of T. erytrae adults. For *T. erytrae* nymphs and eggs (p = 0.190), only

Arusha had positive values, with a mean of 2.508 and a maximum of 43.



Figure 6. Graph of ACP (Adults and Nymphs) and ACT (Adults and Eggs/Nymphs) against regions

An ANOVA test demonstrated that the regions differed significantly in their abundance of *Diaphorina citri*

Kuwayama adults and *Diaphorina citri* Kuwayama nymphs. However, there were no significant differences among the regions for *T. erytrae* adults or *T. erytrae* nymphs and eggs. The Kruskal-Wallis test, a non-parametric alternative to ANOVA, also confirmed the significant differences in *Diaphorina citri* Kuwayama adult abundance among the regions (p-value = 0.0004). To identify which regions differed significantly from each other, a Tukey's HSD post-hoc test was conducted. The results indicated that Tanga had a significantly higher *Diaphorina citri* Kuwayama adult abundance than Arusha

(p-value = 0.0102), Morogoro (p-value = 0.0153), and Ruvuma (p-value = 0.0036). However, no significant differences existed between Arusha and Morogoro, Arusha and Ruvuma, or Morogoro and Ruvuma.

Influence of altitude on psyllid species abundance

The abundance of *Diaphorina citri* Kuwayama nymphs, adults, *T. erytrae* nymphs/eggs, and adults was regressed on altitude using linear regression models. The regression coefficients are presented below in Table 3.

Character	Intercept	Altitude coefficient
ACP Nymphs Total	0.364	-0.0004***
ACP Adults Total	2.586	-0.0028***
ACT Nymphs and Eggs	-1.083	0.0031***
ACT Adults Total	-0.081	0.0002*

We found that altitude notably impacts the prevalence of *Diaphorina citri* Kuwayama nymphs, adults, *T. erytrae* nymphs/eggs, and adults. The abundance of *Diaphorina citri* Kuwayama (both nymphs and adults) was lower at higher altitudes, as indicated by significant negative

coefficients. Conversely, *T. erytrae* (both nymphs/eggs and adults) exhibited a significant increase in abundance at higher altitudes, as indicated by positive coefficients (Figure 7).



Figure 7. Distribution of altitude for citrus psyllids species

The influence of temperature on Psyllid abundance

A correlation analysis was performed to investigate the link between temperature and *Diaphorina citri* Kuwayama abundance (Table 4). The findings indicated a weak positive correlation between these two variables (r = 0.135), suggesting that higher temperatures were associated with slightly higher *Diaphorina citri* Kuwayama abundance. However, the relationship between the two factors was not particularly strong (Figure 8).



Figure 8. A scatter plot of total ACP adults and Nymphs against temperature

Correlation analysis investigated the link between temperature and *T. erytrae* abundance. The findings indicated a moderate negative correlation (r = -0.278)

between the two variables, indicating that higher temperatures were moderately linked to lower *T. erytrae* abundance (Figure 9).



Figure 9. A scatter plot of total ACT adults and Eggs/Nymphs against temperature

The influence of varieties on citrus psyllid abundance An ANOVA was conducted to assess the impact of different citrus varieties on the abundance of *Diaphorina citri Kuwayama*. The findings indicated a significant effect of variety on *Diaphorina citri Kuwayama* abundance (p < 0.001). Furthermore, Fisher's exact test using simulation suggested a significant relationship between the occurrence of *Diaphorina citri Kuwayama* and the various citrus varieties (p<0.046). Additional analysis was performed to identify the most preferred citrus variety by *Diaphorina citri Kuwayama*. The Late Valencia (Kitenesi) variety had an average of 9.63 *Diaphorina citri Kuwayama* counts per tree, the highest average compared to the other studied varieties (Table 5).

Table 5. The mean <i>Diaphorina citri</i> Kuwayama counts per tree on varieties.				
S.N	Variety	Mean D. citri number per tree	S.E	
1	Bitijuma	0.0	0	
2	Jaffa (Shamoti)	0.0	0	
3	Late Valencia (Kitenesi)	9.634	2.815	
4	Local variety	0.0	0	
5	Matombo sweet	0.926	0.544	
6	Mediterranean (Nairobi)	0.0	0	
7	Valencia (Msasa)	0.328	0.123	
8	Washington navel (Kitovu)	0.0	0	

An analysis was conducted to assess the impact of flush intensity on citrus psyllid count. The findings indicated that trees with high flush intensity exhibited a comparatively greater number of citrus psyllid counts (Figure 10). Conversely, trees with low flush intensity had fewer citrus psyllids.

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Figure 10. The relationship between Diaphorina citri Kuwayama and the intensity of new flushes

DISCUSSION

This study unveils substantial geographical disparities in the presence of Diaphorina citri Kuwayama and Trioza erytreae within Tanzania. The presence of Diaphorina citri Kuwayama in the Tanga region was first recorded in this study, representing the initial documentation of its occurrence in this region. Conversely, T. erytrae was confined only to the Arusha region. Various factors contribute to the spread of these insect species across different regions. Among the contributing factors is the potential transportation of citrus plants or infested plant parts, which can serve as vectors for disseminating these insects (Wu et al., 2015). Moreover, weather conditions, especially humidity levels, play a crucial role. Elevated humidity levels can significantly impact the survival and reproductive rates of D. citri while rendering citrus plants more susceptible to infection by Candidatus Liberibacter spp., the pathogens responsible for causing HLB disease (Hall & Hentz, 2011; Yang et al., 2006). These populations depend on many factors, such as the weather (Aidoo et al., 2022; Wang et al., 2020), the number of suitable host plants (Johnston et al., 2019), farming methods, and pest control methods (Kuhns et al., 2016;

Pluke et al., 2008). Notably, the observed distribution pattern underscores the wider geographical expanse of D. citri in Tanzania compared to T. erytrae. These findings have valuable implications for monitoring and managing psyllid populations by shedding light on their spatial dynamics (Grafton-Cardwell et al., 2013; Miranda et al., 2018). This study has yielded crucial insights into the spatial distribution and population density dynamics of D. citri and T. erytrae across different Tanzanian regions. The results highlight that D. citri predominantly thrives in the Tanga region, exhibiting a notably higher population density than the surveyed regions, as visually depicted in Figure 2.5. It is noteworthy that T. erytrae was exclusively detected in the Arusha region. Aidoo et al. (2019) have reported more T. erytrae populations densities in shaded than unshaded trees. The pronounced variations in psyllid abundance observed among distinct regions within the country underscore the localized nature of these infestations. This highlights the need for region-specific management strategies tailored to each locality's unique dynamics. Vikram Singh and Yadav (2018) highlighted that environmental factors, citrus farming methods, and host plant availability impact the varying abundance of D.

citri in different regions. The elevated mean and maximum D. citri count in Tanga suggest that this region may provide a more conducive climate or ecological environment for D. citri populations. Additionally, Beloti et al. (2013) explained that factors like proximity to infested areas, the density of citrus orchards, and the citrus cultivars grown in the area can affect psyllid abundance. The linear regression models employed in this study have yielded compelling evidence of altitude's substantial influence on both D. citri and T. erytrae populations. Shimwela et al. (2016) reported that D. citri nymphs and adults exhibit a negative relationship with altitude, indicating a decrease in their abundance with increasing elevation. Conversely, T. erytrae nymphs/eggs and adults display a positive relationship with altitude, suggesting an increase in their abundance at higher altitudes, as observed in the study by Shimwela et al. (2016). These distinct population patterns can be attributed to factors such as climate preferences, host plant availability, and geographical features. D. citri, a species adapted to tropical and subtropical climates, may prefer lower altitudes characterized by warmer temperatures and ample host plant availability. According to Jenkins et al. (2015), the associated temperature drop could potentially hinder D. citri populations by having an impact on a variety of crucial aspects, such as development, survival, reproduction, or dispersal. On the other hand, according to Aidoo et al. (2021), T. erytrae, which thrives in cooler environments, may find higher altitudes more favourable for their growth, feeding, and oviposition. This emphasizes the crucial role of altitude in developing integrated pest management plans for citrus psyllids, especially in areas with diverse altitude levels. This newfound knowledge equips farmers and stakeholders in the citrus industry with valuable information to assess the potential risks of psyllid infestations across different altitudinal zones and to implement tailored control measures accordingly. The correlation analysis performed in this study showed intricate relationships between temperature and citrus psyllid abundance. We identified a weak positive correlation between temperature and D. *citri* abundance, indicating that higher temperatures may be linked to increased populations of D. citri. However, it is essential to note that the correlation coefficient underscores that temperature alone does not strongly predict D. citri abundance. This aligns with previous research highlighting that, while influential, temperature is merely one of several factors ruling citrus psyllid population dynamics. For instance, Chong et al. (2010) reported a positive correlation between temperature and psyllid density in young citrus groves, while another study by Tomaseto (2016) suggested that temperature impacts the development and activity of D. citri. A moderately negative correlation existed between temperature and T. erytrae abundance, suggesting a potential association between higher temperatures and reduced T. erytrae populations. While studying similar psyllid species can provide some insights (Zavala-Zapata et al., 2022; Tsai et

al., 2002), a more accurate understanding would be gained by investigating the impact of climate changes on T. erytrae in Tanzania. This investigation aims to understand how temperature influences the population growth dynamics of T. erytrae in Tanzania, contributing to the broader understanding of climate effects on psyllid populations. Additionally, our study unveiled significant disparities in D. citri abundance and occurrence across various citrus varieties. This aligns with prior investigations in other countries that have reported varying susceptibility or attractiveness of citrus varieties to D. citri (Eduardo et al., 2022; Nehru et al., 2004). Notably, the Late Valencia (Kitenesi) variety exhibited the highest mean number of D. citri per tree, suggesting it is the preferred choice for the pest. Additionally, its commercial prominence in Tanzania further increases its exposure and accessibility to D. citri. On the other hand, Bitijuma, Jaffa (Shamoti), Local the varieties. Mediterranean (Nairobi), and Washington navel (Kitovu) varieties displayed zero D. citri counts per tree, suggesting potential resistance or repellence to the pest. The Matombo sweet and Valencia (Msasa) varieties exhibited low mean numbers of D. citri per tree, indicating a moderate level of susceptibility or attractiveness to the pest, possibly due to specific characteristics making them less favourable or accessible to D. citri than the Late Valencia (Kitenesi) variety. These findings hold significant implications for the management of Huanglongbing (HLB) and its vector in Tanzania and other citrus-producing regions in East Africa. One key strategy to bolster HLB control efforts is to reduce the population and activity of D. citri (Li et al., 2020). A promising avenue to pursue in this regard is the selection and cultivation of citrus varieties that display resistance to this pest, as proposed by Borgoni et al. in 2014. This research identified five citrus varieties that consistently show zero D. citri counts per tree. This discovery holds great promise for advancing HLB-resistant citrus breeding programs and integrated pest management practices in Tanzania and neighbouring East African countries (Table 6). These varieties can serve as valuable candidates for further investigation and development (Chen & Stelinski, 2017; Eduardo et al., 2022). Still, it's important to stress the need for more research to confirm whether these identified varieties are resistant or repellent to D. citri in a range of environmental conditions. Based on our research, we have found that the presence of new flushes greatly influences the occurrence and abundance of citrus psyllids. Specifically, citrus plants that exhibit moderate to high intensity of new flushes had a significantly higher number of D. citri per plant, as demonstrated in Figure 2.7. Similar studies were conducted by Tsai et al. (2002) and Hall et al. (2008), which showed that citrus psyllids abundance correlated with the intensity of new flushes. Adult psyllids are attracted to new flushes due to the yellowish color they produce (Wu et al., 2015). In addition, new flushes release volatile organic compounds (VOCs) that attract D. citri from a distance (Zhao *et al.*, 2013). Therefore, *D. citri* has developed strategies to deal with the changing availability of new flushes. For example, during periods of low flush availability, *D. citri* enters a state of

reproductive diapause (temporary cessation of reproduction) until new flushes appear, at which point they resume reproduction (Britt *et al.*, 2020).

Table 4. Overview of successful measures for managing various pests and diseases, recommended for other parts of
Africa cultivating citrus and fruit trees.

Management Practice	Description	Applicability in Africa		
Biopesticides	Use of natural organisms like <i>Beauveria bassiana</i> and botanical pesticides (Ramírez-Godoy <i>et al.</i> , 2018).	Highly recommended due to environmental sustainability and effectiveness.		
Biodiversity Management	Encouraging natural predators and functional biodiversity through organic farming practices (Rugno <i>et al.</i> , 2021)	Suitable for organic orchards; promotes resistance to pest proliferation.		
Cultural Practices	Includes sanitation, pruning, mulching, and weed management (Lucas, 2011).	Essential for integrated pest management; applicable across various regions.		
Chemical Control	Judicious use of insecticides and mineral oils (Leong <i>et al.</i> , 2022).	To be used cautiously; recommended in combination with other IPM strategies.		
Biological Control	Utilization of natural enemies and biocontrol agents (Rugno <i>et al.</i> , 2021).	Effective and environmentally friendly; encourages local biodiversity.		
Quarantine Measures	Inspection and certification to prevent the spread of pests (Leong <i>et al.</i> , 2022).	Critical for maintaining market access and preventing regional spread.		
Integrated Pest Management (IPM)	Combination of various control measures for long-term sustainability (Leong <i>et al.</i> , 2022).	Recommended as a holistic approach to pest management.		

CONCLUSION

This research provides valuable insights into the geographical disparities and distribution patterns of *D. citri* and *T. erytreae* within Tanzania. Our study underscores the predominance of *D. citri* in the Tanga and Morogoro regions, with the Tanga region serving as a newly documented area of infestation. Conversely, *T. erytrae* is exclusively confined to the Arusha region. These observed variations in psyllid abundance across regions underscore the localized nature of these infestations and underscore the critical need for tailoring management strategies to the specific characteristics of each region (Table 4).

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REFERENCES

- Aidoo, O. F., Souza, P. G. C., da Silva, R. S., Santana, P. A., Picanço, M. C., Kyerematen, R., Sètamou, M., Ekesi, S., & Borgemeister, C. (2022). Climate-induced range shifts of invasive species (*Diaphorina citri* Kuwayama). *Pest Management Science* 78(6): 2534 2549. doi: <u>10.1002/ps.6886</u>
- Aidoo, O. F., Tanga, C. M., Mohamed, S. A., Khamis, F. M., Opisa, S., Rasowo, B. A., Kimemia, J. W., Ambajo, J., Sétamou, M., Ekesi, S., & Borgemeister, C. (2021). The African citrus triozid *Trioza erytreae* Del Guercio (Hemiptera: Triozidae): temporal dynamics and susceptibility to entomopathogenic

fungi in East Africa. International Journal of Tropical Insect Science 41(1): 563 – 573. doi: 10.1007/s42690-020-00241-5

- Aidoo, O. F., Tanga, C. M., Mohamed, S. A., Rasowo, B.
 A., Khamis, F. M., Rwomushana, I., ... & Borgemeister, C. (2019). Distribution, degree of damage and risk of spread of *Trioza erytreae* (Hemiptera: Triozidae) in Kenya. *Journal of Applied Entomology*, 143(8), 822-833. doi: 10.1111/jen.12668
- Aminu, F. O., Balogun, E. O. S., & Ojo, O. O. (2020). Analysis of post-harvest losses in orange in Lagos metropolitan markets, Nigeria. NIGERIAN JOURNAL OF HORTICULTURAL SCIENCE (NJHS), 143.
- Andrade-Cuvi, M. J., Montalvo, C., & Moreno-Guerrero, C. (2023). Vegetable and fruit consumption during the COVID-19 lockdown: eating habits in Ecuador. Agricultural and Food Economics, 11(1), 37. doi: 10.1186/s40100-023-00260-z
- Beloti, V. H., Rugno, G. R., Felippe, M. R., Carmo-Uehara, A. Do, Garbim, L. F., Godoy, W. A. C., & Yamamoto, P. T. (2013). Population dynamics of *Diaphorina citri* kuwayama (hemiptera: Liviidae) in orchards of "valencia" orange, "ponkan" mandarin and "murcott" tangor trees. *Florida Entomologist* 96(1): 173 179. doi: 10.1653/024.096.0123
- Borgoni, P. C., Vendramim, J. D., Lourencão, A. L., & Machado, M. A. (2014). Resistance of Citrus and Related Genera to *Diaphorina citri* Kuwayama (Hemiptera: Liviidae). *Neotropical Entomology* 43(5): 465–469. doi: 10.1007/s13744-014-0230-0

- Boykin, L. M., De Barro, P., Hall, D. G., Hunter, W. B., McKenzie, C. L., Powell, C. A., & Shatters, R. G. (2012). Overview of worldwide diversity of *Diaphorina citri* Kuwayama mitochondrial cytochrome oxidase 1 haplotypes: Two Old World lineages and a New World invasion. *Bulletin of Entomological Research* 102(5): 573 – 582. doi: 10.1017/S0007485312000181
- Britt, K., Gebben, S., Levy, A., Al Rwahnih, M., & Batuman, O. (2020). The detection and surveillance of Asian Citrus Psyllid (*Diaphorina citri* Kuwayama)—associated viruses in Florida citrus groves. *Frontiers in Plant Science*, 10, 1687.
- Chen, X. D., & Stelinski, L. L. (2017). Resistance management for Asian citrus psyllid, *Diaphorina citri* Kuwayama, in Florida. *Insects*, 8(3), 103. doi: <u>10.3390/insects8030103</u>
- Chong, J. H., Roda, A. L., & Mannion, C. M. (2010). Density and natural enemies of the Asian citrus psyllid, *Diaphorina citri* (Hemiptera: Psyllidae), in the residential landscape of Southern Florida. *Journal of Agricultural and Urban Entomology* 27(1): 33 – 49. doi: 10.3954/11-05.1
- De Oliveira, J. M. C., do Nascimento, A. S., de Miranda, S. H. G., Barbosa, C. de J., & Laranjeiras, F. F. (2013). EStimating the economic impact of an eventual introduction of huanglongbing (HLB) in the state of Bahia, Brazil. *Revista Brasileira de Fruticultura* 35(3): 755 – 762. doi: <u>10.1590/S0100-29452013000300012</u>
- Economic and Social Research Foundation (ESRF). (2021). Citrus in Tanzania [PDF]. <u>https://esrf.or.tz/wpcontent/uploads/2021/05/Tanzania-Citrus-Brief.pdf</u>
- Eduardo, W. I., Miranda, M. P., Volpe, H. X. L., Garcia, R. B., Girardi, E. A., Alquezar, B., Ruiz, A. E., & Peña, L. (2022). Resistance of true citrus species to *Diaphorina citri* Kuwayama. *Pest Management Science* 78(11): 4783–4792. doi: <u>10.1002/ps.7098</u>
- FAO (2021). Citrus fruit. Fresh and processed- statistical bulletin 2020. Trade and markets division, [http://www.fao.org/3/a-i8092e.pdf] site visited on 20/5/2023.
- FAOSTAT (2020). Food and agriculture data. [https://www.fao.org/faostat/en/#data] site visited on 20/2/2023.
- Fujiwara, K., Uechi, N., Shimizu, Y., Toda, S., Inoue, H., Yamaguchi, T., ... & Fujikawa, T. (2017). Effective molecular detection of *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) in bulk insect samples from sticky traps. *Journal of Applied Entomology*, 141(1-2), 61-66. doi: <u>10.1111/jen.12315</u>
- Garnier, M., Danel, N., & Bové, J. M. (1984a). Aetiology of citrus greening disease. Annales de l'Institut Pasteur Microbiology 135(1): 169–171. doi: 10.1016/S0769-2609(84)80075-7
- Garnier, M., Danel, N., & Bové, J. M. (1984b). The Greening Organism is a Gram-Negative Bacterium. *International Organization of Citrus Virologists Conference Proceedings*. pp. 115 – 124. doi: 10.5070/c59277j4jm

- Grafton-Cardwell, E. E., Stelinski, L. L., & Stansly, P. A. (2013). Biology and management of Asian citrus psyllid, vector of the huanglongbing pathogens. In *Annual Review of Entomology* 58: 413 432. doi: 10.1146/annurev-ento-120811-153542
- Hall, D. G., & Hentz, M. G. (2011). Seasonal flight activity by the Asian citrus psyllid in east central Florida. *Entomologia Experimentalis et Applicata* 139(1): 75 – 88. doi: <u>10.1111/j.1570-</u> <u>7458.2011.01108.x</u>
- Hall, D. G., & Hentz, M. G. (2016). An Evaluation of Plant Genotypes for Rearing Asian Citrus Psyllid (Hemiptera: Liviidae). *Florida Entomologist* 99(3): 471–480. doi: <u>10.1653/024.099.0320</u>
- Hall, D. G., Hentz, M. G., & Adair Jr, R. C. (2008). Population ecology and phenology of *Diaphorina citri* (Hemiptera: Psyllidae) in two Florida citrus groves. *Environmental Entomology*, 37(4), 914-924. doi: 10.1603/0046-225X(2008)37[914:PEAPOD]2.0.CO;2

Hall, D. G., Richardson, M. L., Ammar, E. D., & Halbert,

- Fran, D. G., Kichardson, M. L., Annnar, E. D., & Halbert, S. E. (2013). Asian citrus psyllid, *Diaphorina citri*, vector of citrus huanglongbing disease. In *Entomologia Experimentalis et Applicata* 146(2): 207–223. doi: <u>10.1111/eea.12025</u>
- Hall, D., Ammar, E. D., Bowman, K., & Stover, E. (2018). Epifluorescence and stereomicroscopy of trichomes associated with resistant and susceptible host plant genotypes of the asian citrus psyllid (Hemiptera: Liviidae), Vector of citrus greening disease bacterium. *Journal of Microscopy and Ultrastructure* 6(1): 1 – 56. doi: 10.4103/jmau.jmau_9_18
- Jenkins, D. A., Hall, D. G., & Goenaga, R. (2015). Diaphorina citri (Hemiptera: Liviidae) abundance in puerto rico declines with elevation. Journal of Economic Entomology 108(1): 252 – 258. doi: 10.1093/jee/tou050
- Johnston, N., Stansly, P. A., & Stelinski, L. L. (2019). Secondary hosts of the Asian citrus psyllid, *Diaphorina citri* Kuwayama: Survivorship and preference. *Journal of Applied Entomology* 143(9): 921 – 928. doi: <u>10.1111/jen.12673</u>
- Kuhns, E. H., Martini, X., Hoyte, A., & Stelinski, L. L. (2016). Repellent activity of botanical oils against Asian citrus psyllid, *Diaphorina citri* (hemiptera: Liviidae). *Insects* 7(3): 1 35. doi: 10.3390/insects7030035
- Lahlali, R., Jaouad, M., Moinina, A., Mokrini, F., & Belabess, Z. (2021). Farmers' knowledge, perceptions, and farm-level management practices of citrus pests and diseases in Morocco. Journal of Plant Diseases and Protection, 128(5), 1213-1226. doi: <u>10.1007/s41348-021-00479-2</u>
- Lallemand, J., Fos, A., & Bové, J. M. (1986). Transmission de la bactérie associée à la forme africaine de la maladie du" greening" par le psylle asiatique *Diaphorina citri* Kuwayama. *Fruits*, 41(5), 341-343.
- Leong, S. S., Leong, S. C., & Beattie, G. A. (2022). Integrated pest management strategies for Asian

citrus psyllid *Diaphorina citri* Kuwayama (Hemiptera: Psyllidae) and huanglongbing in citrus for Sarawak, East Malaysia, Borneo. Insects, 13(10), 960. doi: <u>10.3390/insects13100960</u>

- Li, S., Wu, F., Duan, Y., Singerman, A., & Guan, Z. (2020). Citrus greening: Management strategies and their economic impact. *HortScience* 55(5): 604–612. doi: 10.21273/HORTSCI14696-19
- Liu, S., Lou, Y., Li, Y., Zhang, J., Li, P., Yang, B., & Gu, Q. (2022). Review of phytochemical and nutritional characteristics and food applications of Citrus L. fruits. In *Frontiers in Nutrition* 9: 1 – 16. doi: 10.3389/fnut.2022.968604
- Lucas, J. A. (2011). Advances in plant disease and pest management. The Journal of Agricultural Science, 149(S1), 91-114. doi: <u>10.1017/s0021859610000997</u>
- Makorere, R. (2014). an Exploration of Factors Affecting Development of Citrus Industry in Tanzania: Empirical Evidence From Muheza District, Tanga Region. *International Journal of Food and Agricultural Economics* 2(2): 135–154.
- Miranda, M. P., dos Santos, F. L., Bassanezi, R. B., Montesino, L. H., Barbosa, J. C., & Sétamou, M. (2018). Monitoring methods for *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) on citrus groves with different insecticide application programmes. *Journal of Applied Entomology* 142: 1–2. doi: 10.1111/jen.12412
- Moreb, N. A., Albandary, A., Jaiswal, S., & Jaiswal, A. K. (2021). Fruits and vegetables in the management of underlying conditions for COVID-19 high-risk groups. *Foods*, *10*(2), 389. doi: 10.3390/foods10020389
- Narouei-Khandan, H. A., Halbert, S. E., Worner, S. P., & van Bruggen, A. H. (2016). Global climate suitability of citrus huanglongbing and its vector, the Asian citrus psyllid, using two correlative species distribution modeling approaches, with emphasis on the USA. *European Journal of Plant Pathology*, *144*, 655-670. doi: <u>10.1007/s10658-015-0804-7</u>
- National Academies of Sciences, Engineering, and Medicine (2018). A Review of the Citrus Greening Research and Development Efforts Supported by the Citrus Research and Development Foundation: *Fighting a Ravaging Disease*. The National Academies Press, Washington, DC, USA, 288 pp. doi: 10.17226/25026
- Nehru, R. K., Bhagat, K. C., & Koul, V. K. (2004). Influence of citrus species on the development of *Diaphorina citri* Kuwayama. In *Annals of Plant Protention Sciences* 12(2): 425–438.
- Pluke, R. W. H., Qureshi, J. A., & Stansly, P. A. (2008). Citrus flushing patterns, *Diaphorina citri* (Hemiptera: Psyllidae) populations and parasitism by *Tamarixia radiata* (Hymenoptera: Eulophidae) in Puerto Rico. *Florida Entomologist* 91(1): 36 – 42. doi: <u>10.1653/0015-</u> <u>4040(2008)091[0036:CFPDCH]2.0.CO;2</u>
- Ramírez-Godoy, A., del Pilar Vera-Hoyos, M., Jiménez-Beltrán, N., & Restrepo-Díaz, H. (2018). Evaluation

of three commercial biopesticides for the reduction of *Diaphorina citri* (Hemiptera: Liviidae) populations. *HortScience*, *53*(10), 1453-1460. doi: <u>10.21273/HORTSCI13349-18</u>

- Rasowo, B. A., Khamis, F. M., Mohamed, S. A., Ajene, I.
 J., Aidoo, O. F., Ombura, L., ... & Borgemeister, C.
 (2019). African citrus greening disease in East Africa: incidence, severity, and distribution patterns. Journal of Economic Entomology, 112(5), 2389-2397. doi: <u>10.1093/jee/toz167</u>
- Reportlinker. (2023). Forecast: Citrus production in the World 2022 - 2026. [https://www.Reportlinker.com/dataset/e130a54a610e47bdb73fae99d2 6bc1b7b7d0676]site visited on 20/3/2023.
- Rogers, M. E., & Stansly, P. A. (2007). Biology and management of the asian citrus psyllid, *Diaphorina citri* Kuwayama, in Florida Citrus. [https://swfrec.ifas.ufl.edu/hlb/databas e/pdf/00003031.pdf] site visited on 22/3/2023. doi: 10.32473/edis-in668-2006
- Rugno, G. R., Cuervo, J. G. B., Garcia, A. G., Qureshi, J., & Yamamoto, P. T. (2021). Abundance and diversity of lacewings in grower operated organic and conventional pest management programs for *Diaphorina citri* (Hemiptera: Liviidae). Crop Protection, 146, 105682. doi: 10.1016/j.cropro.2021.105682
- Rwomushana, I., Khamis, F. M., Grout, T. G., Mohamed, S. A., Sétamou, M., Borgemeister, C., Heya, H. M., Tanga, C. M., Nderitu, P. W., Seguni, Z. S., Materu, C. L., & Ekesi, S. (2017). Detection of *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) in Kenya and potential implication for the spread of Huanglongbing disease in East Africa. *Biological Invasions* 19(10): 2777–2787. doi: 10.1007/s10530-017-1502-5
- Shimwela, M. M., Narouei-Khandan, H. A., Halbert, S. E., Keremane, M. L., Minsavage, G. V., Timilsina, S., Massawe, D. P., Jones, J. B., & van Bruggen, A. H. C. (2016). First occurrence of *Diaphorina citri* in East Africa, characterization of the Ca. Liberibacter species causing huanglongbing (HLB) in Tanzania, and potential further spread of *Diaphorina citri* and HLB in Africa and Europe. *European Journal of Plant Pathology* 146(2): 349–368. doi: 10.1007/s10658-016-0921-y
- Singerman, A., & Useche, P. (2019). The Role of Strategic Uncertainty in Area-wide Pest Management Decisions of Florida Citrus Growers. *American Journal of Agricultural Economics* 101(4): 991 – 1011. doi: 10.1093/ajae/aaz006
- Swai, I. S. (1985, November). Citrus diseases in Tanzania. In XII African Symposium on Horticultural Crops 218 (pp. 329-332).
- Tomaseto, A. F. (2016, August 2). Effect of temperature on *Diaphorina citri* (Hemiptera: Liviidae) take-off. [https://doi.org/10.1603/ice.2016.114432] site visited on 20/3/2023.
- Tsai, J. H., Wang, J. J., & Liu, Y. H. (2002). Seasonal abundance of the Asian citrus psyllid, *Diaphorina citri* (Homoptera: Psyllidae) in Southern Florida.

African J. Trop. Entomol. Res. 3(2):52-64

Florida Entomologist, 85(3): 446 – 451. doi: <u>10.1653/0015-</u> 4040(2002)085[0446:SAOTAC]2.0.CO;2

- URT (2021). National Sample Census of Agriculture 2019/20. National Bureau of Statistics, Dodoma. 931pp.
- Vikram Singh, N., & Yadav, G. S. (2018). Bio-ecology and Management of Citrus psylla, *Diaphorina citri* Kuwayama on Citrus - A Review. *International Journal of Current Microbiology and Applied Sciences* 7(04): 3091 – 3107. doi: 10.20546/ijcmas.2018.704.351
- Wang, R., Yang, H., Wang, M., Zhang, Z., Huang, T., Wen, G., & Li, Q. (2020). Predictions of potential geographical distribution of *Diaphorina citri* (Kuwayama) in China under climate change scenarios. *Scientific Reports* 10(1): 1 – 9. doi: 10.1038/s41598-020-66274-5
- Wangithi, C. M. (2019). Evaluation of the magnitude of Citus yield losses due to African Citrus Triozid, False Codling Moth, the Greening Disease and other pests of economic importance in Kenya. doi: 10.7176/jesd/10-20-11
- Wu, F., Cen, Y., Deng, X., Chen, J., Xia, Y., & Liang, G.

(2015). Movement of *Diaphorina citri* (Hemiptera: Liviidae) adults between huanglongbing-infected and healthy citrus. *Florida Entomologist*, *98*(2), 410-416. doi: 10.1653/024.098.0203

- Yang, Y., Huang, M., Andrew, G., Beattie, C., Xia, Y., Ouyang, G., & Xiong, J. (2006). Distribution, biology, ecology and control of the psyllid *Diaphorina citri* Kuwayama, a major pest of citrus: A status report for China. *International Journal of Pest Management* 52(4); 343 – 352. doi: 10.1080/09670870600872994
- Zavala-Zapata, V., Lázaro-Dzul, M. O., Sánchez-Borja, M., Vargas-Tovar, J. A., Álvarez-Ramos, R., & Azuara-Domínguez, A. (2022). Abundance of *Diaphorina citri* kuwayama associated with temperature and precipitation at Tamaulipas, Mexico. *Southwestern Entomologist* 47(3): 713 – 722. doi: 10.3958/059.047.0321
- Zhao, J. P., Wang, H. T., Zeng, X. N., & Xue, P. P. (2013). Differences in selection behaviors and chemical cues of adult Asian citrus psyllids, *Diaphorina citri*, on healthy and huanglongbing-infected young shoots of citrus plants. *Journal of Agricultural Science*, 5(9), 83. doi: 10.5539/jas.v5n9p83